

Effect of lead on male gonadal activity in Albino Rats

Biswas NM¹ Ghosh P²

¹Professor of Physiology, Kathmandu Medical College, Kathmandu, ²Senior teacher of Biology, Sahid Rameswar Vidyamandir, Calcutta, India.

Abstract

Lead poisoning often prevails in children and industrial workers. The present study was undertaken to evaluate the effects of lead acetate on steroidogenic functions of testis, serum levels of gonadotrophins and testosterone in albino rats. Testicular steroidogenic activity was evaluated by measuring the activities of two steroidogenic key enzymes, Δ^5 - 3β hydroxysteroid dehydrogenase (Δ^5 - 3β -HSD) and 17β -hydroxysteroid dehydrogenase (17β -HSD). Administration of lead acetate at a dose of 8mg/kg body weight for 14 days lowered the weights of testes and accessory sex organs, and decreased testicular Δ^5 - 3β -HSD and 17β -HSD activities and serum levels of FSH, LH and testosterone but 7 days of lead acetate administration showed no effect on the above parameters. This report is perhaps the first evidence to show that lead exerts some deleterious effects on testicular steroidogenesis indirectly by decreasing serum levels of gonadotropins.

Keywords: Lead, male gonad, gonadotropins, Albino Rat

Lead is a heavy soft metal, occurs in nature as an oxide or salts. Chronic lead poisoning is commonly seen in young children from sucking lead paint or lead toys, in workers engaged in printing, paint and petroleum industries. Effects of chronic lead poisoning are the disorders in gastrointestinal and haemopoietic system, and muscular weakness leading to paralysis.¹

Literature survey so far available indicates that lead treatment causes anemia.² Testicular histochemical changes and spermatogenic inhibition have also been observed after lead administration in rats.^{3,4}

Though the lead administration results the fall in the serum level of testosterone in rats,^{5,6} the effect of lead salt on testicular steroidogenesis and its mechanism of action on the male gonads have not been studied.

Therefore, the present study has been undertaken on albino rats to investigate the effects of lead acetate on testicular steroidogenic enzymes and serum levels of gonadotropins and testosterone.

Materials and methods

The present study was conducted (with 32 mature male albino rats) in the laboratory of the department of physiology, Calcutta University, Calcutta, India. The animals were of Wistar strain, weighing 150 – 170gm, divided equally into two groups and maintained under standard laboratory conditions (14 h light 10h darkness; at $28 \pm 4^\circ\text{C}$) with standard animal diet and water available ad libitum. Lead

acetate was purchased from E, Merck Chemical Company (Bombay, India). It was dissolved in sterile distilled water. Eight animals of one group received 1ml sterile distilled water kg^{-1} for 7 days and eight animals of other group received the same vehicle for 14 days. The remaining eight animals of each group were injected intraperitoneally with lead acetate at a dose of 8.0mg (1.0ml distilled water), $\text{kg}^{-1} \text{day}^{-1}$ for the same duration as vehicle injected rats. Animals were killed on 8th and 15th day of treatment following protocols and ethical procedure, and their body weights were noted. Blood was collected from dorsal aorta, centrifuged and serum was stored at -20°C for radioimmunoassay. The testes were dissected out and used for enzymatic study.

Measurement of testicular enzymes

One testis from each animal was used for studying the activities of Δ^5 - 3β -hydroxysteroid dehydrogenase (Δ^5 - 3β -HSD) and 17β -hydroxysteroid dehydrogenase (17β -HSD). Testicular Δ^5 - 3β -HSD and 17β -HSD were measured in UV spectrophotometer according to the procedure of Talalay (1962)⁷ and Jarabak et al (1962)⁸, and subsequently modified by Biswas et al (1983; 2003; 2004)⁹⁻¹¹. One unit for both Δ^5 - 3β - and 17β -HSD was defined as the amount causing a change in absorbance of 0.001/min at 340nm.

Correspondence

Prof. Dr. Narendra Mohan Biswas, Msc., Ph.D., D.Sc.
Professor, Dept of Physiology, Kathmandu Medical College (TH),
Sinamangal, KTM

Radioimmunoassay of hormones

Serum FSH and LH were measured according to Moudgal and Madhwaraj (1974)¹² by RIA using reagents supplied by the Rat Pituitary Distribution Program and NIDDK (Bethesda, MD, USA). Pure rat FSH (NIDDK-rFSH¹⁻⁵) were iodinated using the chloramine T (Sigma chemical co., St. Louis, MO, USA) method according to Greenwood et al (1963).¹³ NIDDK anti-rat-FSH-S-II and NIDDK-anti-rat-LH-S-5 were used as antisera. Goat anti rabbit γ -globulin was used as the second antibody. Serum samples were expressed as $\mu\text{g/L}$ of serum. The intraassay variations were 6% and 5% for FSH and LH respectively.

Serum testosterone was assayed according to the procedure of Auletta et al. (1974).¹⁴ Samples were assayed in duplicate. The antiserum to testosterone

was purchased from Endocrine Science (Tarazone, CA, USA) and it had a 44% cross reactivity with 5α -dihydrotestosterone (DHT). The intra assay variation was 6.0%. Reported values are the sum of testosterone and dihydrotestosterone as chromatographic purification of testosterone and dihydrotestosterone are not possible.

Statistical Analysis for statistical analysis of the data, the two-tailed students' t-test was used. Differences were considered significantly when $p < 0.05$.

Results

Table 1: Changes in body weight and testicular, prostatic and seminal vesicular weight after lead acetate administration at a dose of 8mg/kg body wt' values are mean \pm SEM (n = 8).

Duration of Treatment	Condition	Body weight (g)		Testis	Prostate	Seminal Vesicle
		Initial	Final	(g/kg body wt)	(mg/kg body wt)	(mg/kg body wt)
7 days	Control	150.8 \pm 8.5	161.9 \pm 5.7	24.8 \pm 0.3	845 \pm 47	1235 \pm 110
	Lead	153 \pm 10.3	160.3 \pm 8.5	23.9 \pm 0.5	859 \pm 52	1147 \pm 100
14 days	Control	155.9 \pm 9.2	163.8 \pm 8.2	24.9 \pm 0.8	905 \pm 40	1285 \pm 90
	Lead	156.8 \pm 7.5	168.2 \pm 10.1	20.3 \pm 0.5*	795 \pm 0.38*	930 \pm 70*

P – Values * < 0.05 compared with the corresponding vehicle treated controls.

Table 2: Serum levels of FSH, LH, testosterone and testicular Δ^5 - 3β -HSD, 17β -HSD activities after lead acetate administration at the dose 8mg/kg body wt. Each value represents mean \pm SEM (n=8).

Duration of Treatment	Condition	Serum FSH $\mu\text{g/L}$	Serum LH $\mu\text{g/L}$	Serum Testosterone $\mu\text{g/L}$	Δ^5 - 3β -HSD (unit/mg tissue per h)	17β -HSD (unit/mg tissue per h)
7 days	Control	195.32 \pm 10.28	38.62 \pm 3.39	3.25 \pm 0.19	25.12 \pm 0.152	26.13 \pm 0.35
	Lead	181.49 \pm 9.25	35.91 \pm 2.83	3.41 \pm 0.21	24.41 \pm 0.132	25.49 \pm 0.41
14 days	Control	186.25 \pm 8.39	40.52 \pm 3.28	3.56 \pm 0.20	26.39 \pm 0.141	27.82 \pm 0.58
	Lead	152.38 \pm 10.42*	29.59 \pm 2.62*	2.15 \pm 0.17*	20.09 \pm 0.140*	21.62 \pm 0.43*

P. – Values * < 0.05 compared with the corresponding vehicle treated controls.

Body and organ weights

Lead acetate administration resulted in no significant effect on body weight. Testicular and accessory sex organ weights were decreased remarkably after 14 days of lead administration, but there was no significant effect on the above reproductive organs after lead acetate administration for 7 days in comparison with control animals (Table 1).

Enzymatic study

Testicular (Δ^5 - 3β -HSD) and 17β -HSD activities were decreased remarkably after 14 days of lead acetate administration in respect to control. Seven days of lead administration has no effect on the activities of above steroidogenic enzymes (Table 2).

Hormones

Serum levels of FSH, LH & Testosterone were significantly lowered after lead acetate administration for 14 days but administration of lead for 7 days

resulted in no effect on the serum levels of gonadotropins and testosterone (Table 2).

Discussion

The present study demonstrates that 14 days after administration of lead acetate in rats, there is a significant decrease in the activities of steroidogenic enzymes and fall of serum levels of FSH, LH and testosterone. $\Delta^5-3\beta$ -HSD and 17β -HSD, key steroidogenic enzymes in androgen biosynthesis^{15, 16} are controlled partly by LH¹⁷ and partly by FSH as FSH plays a role in induction of Leydig cell LH receptor.^{18,19} Therefore, the decreased level of testosterone and reduction of the activities of testicular steroidogenic enzymes in lead-treated rats is a reflection of reduced pituitary gonadotropins secretion.²⁰ The mechanism by which the levels of gonadotropins decrease in lead-treated rat can not be determined from the present experiments. But the effect of lead acetate on serum level of testosterone is consistent with others^{5,6}, where lead administration results a fall of serum level of testosterone. Moreover testicular size and weight are normally regulated by fluid secretion from Sertoli cells and the production of sperm in the seminiferous tubules.^{21,22} Reduction of testicular weight less than control values after 14 days of lead administration is in support of degeneration of germ cells and Sertoli cells in lead-treated rats.

In conclusion, our findings suggest that chronic administration of lead to normal mature albino rats can reduce the serum levels of FSH, LH and testosterone which in turn result inhibition of testicular activity and the fall in accessory sex organs weight.

Acknowledgements

We are grateful to Prof. Hemang Dixit, Principal, Col. Shashi Pratap K. C., Sr. Administrator and Prof. Veena Bapat, Head of the Department of Physiology, Kathmandu Medical College, Nepal for constant encouragements for this work. Thank are also due to Mrs. Shanta Karki for her co-operation and computerization of the manuscript.

References

1. Satoskar RS, Bhandarkar, SD, Alnapure SS, Pharmacology and Pharmacotherapeutics, 6th edn, popular Prakashan, Mumbai, India, 1999; 1012 – 1013.
2. Poulos L, Gammaz S, et al. Statistically significant hematopoietic effects of low blood levels. Arch Environ Health 1986; 41: 384 – 6.

3. Chowdhury AR, Rao RV, Gautan AK. Histochemical changes in the tests of lead induced experimental rats. Folia Histochem Cytobiol, 1986; 24: 233 – 7.
4. Guloik ME, Spermatogenesis and maturation of spermatozoa in rats exposed to lead, Ann Acad Med Stetin, 1989; 35:73 – 87.
5. Sokol PZ, Madding CE, Swerdloff PS. Lead toxicity and the hypothalamic- pituitary testicular axis, Biol Reprod, 33: 722 – 728.
6. Sokol PZ. Hormonal effects of lead acetate in the male rat: mechanism of action. Biol Reprod. 1987; 37: 1135 – 1138.
7. Talalay P. Hydroxysteroid dehydrogenase in: Colowick, Kaplan eds. Methods in enzymology. New York: Academic Press, 1962; 5: 512 – 516.
8. Jarabak I, Adams JA, et al. Purification of a 17β - hydroxy steroid dhydrogenase of human placenta and studies on its transhydrogenase function. J Biolchem. 1962; 237: 345 – 357.
9. Biswas NM, Ghosh PK, Neuhaus OW. Effect of α_{2u} - globulin on serum concentration of gonadotropins and testicular activity in oestrogen²⁴ – treated rats. J Endocrinol. 1983; 96: 321 – 327.
10. Biswas NM, Roy Chaudhuri G, et al. Effect of casein diet on gonadotropin releasing hormone antagonist induced changes in adrenal gonadal functions in male rats. Indian J Exp Biol. 2001; 39:1249 – 1253.
11. Biswas NM, Chattopadhyay A, Sarkar M. Effect of gold on testicular steroidogenic and gametogenic functions in immature male albino rats. Life sci. 2004; in Press.
12. Moudgal NR, Madhwa Raj HG, Pituitary gonadotropin. In: Jaffe BM, Behrman HR. eds, Methods of hormone radioimmunoassay. New York: Academic Press, 1974; PP 57 – 85.
13. Greenwood FC, Hunter WM, Glover JS. The preparation of ¹³¹I – labelled human growth hormone of high specific activity. Biochem J. 1963; 89: 144 – 123.
14. Auleta PJ, Caldwell BV, Hamilton G. Androgen, testosterone and dihydrotestosterone. In : Jaffe BM, Behrman HR, eds. Methods of hormone radioimmunoassay. New York: Academic Press, 1974; PP 359 – 370.
15. Maier DM. Species variation in testicular $\Delta^5-3\beta$ -hydroxysteroid dehydrogenase activity: Absence of activity in primate

- Leydig cell. *Endocrinology* 1965; 76: 463 – 469.
16. Baillie AH, Mack WS. Hydroxysteroid dehydrogenase in normal and abnormal human testes. *J. Endocrinol.* 1966; 35: 239 – 248.
 17. Ramirez DV, McCann SM. Comparison of the regulation of LH secretion in immature and adult rats. *Endocrinology*, 1963; 72: 452 – 464.
 18. Swerdloff RS, Walsh PC, et al. Serum LH and FSH during sexual maturation in the male rat: Effect of castration and cryptorchidism. *Endocrinology*, 1971: 88: 120 – 128.
 19. Odell WD, Swerdloff RS, et al. FSH induction of sensitivity to LH: one cause of sexual maturation in the male rat. *Endocrinology*, 1973; 92: 160 – 165.
 20. Biswas NM, Ghosh PK, et al. Effect of thyroidectomy, and thyroxine and α_{2u} - globulin replacement therapy on testicular steroidogenic and gametogenic activities in rats. *J Endocrinol.* 1994; 140: 343 – 347.
 21. Waites GMH Gladwell RT. Physiological significance of fluid secretion in the testis and blood testis barrier. *Physiol Rev.* 1982; 62: 624 – 671.
 22. Aman RP. Sperm production rates. In: Johnson AD, Gomes WR, Van Damark NL. eds. *The testis*, Vol. 1. New York: Academic Press, 1970; PP 433 – 482.